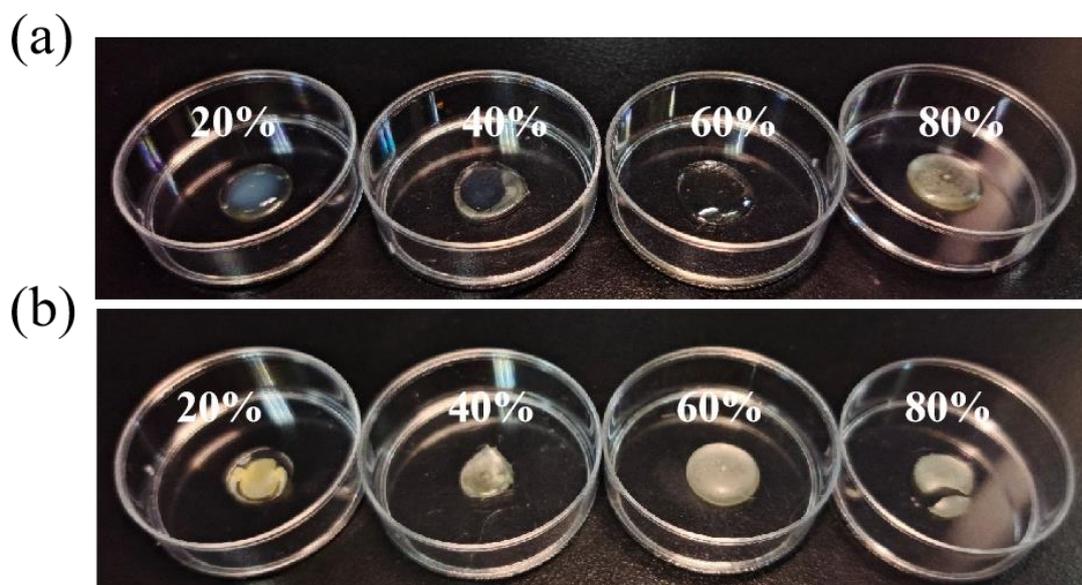


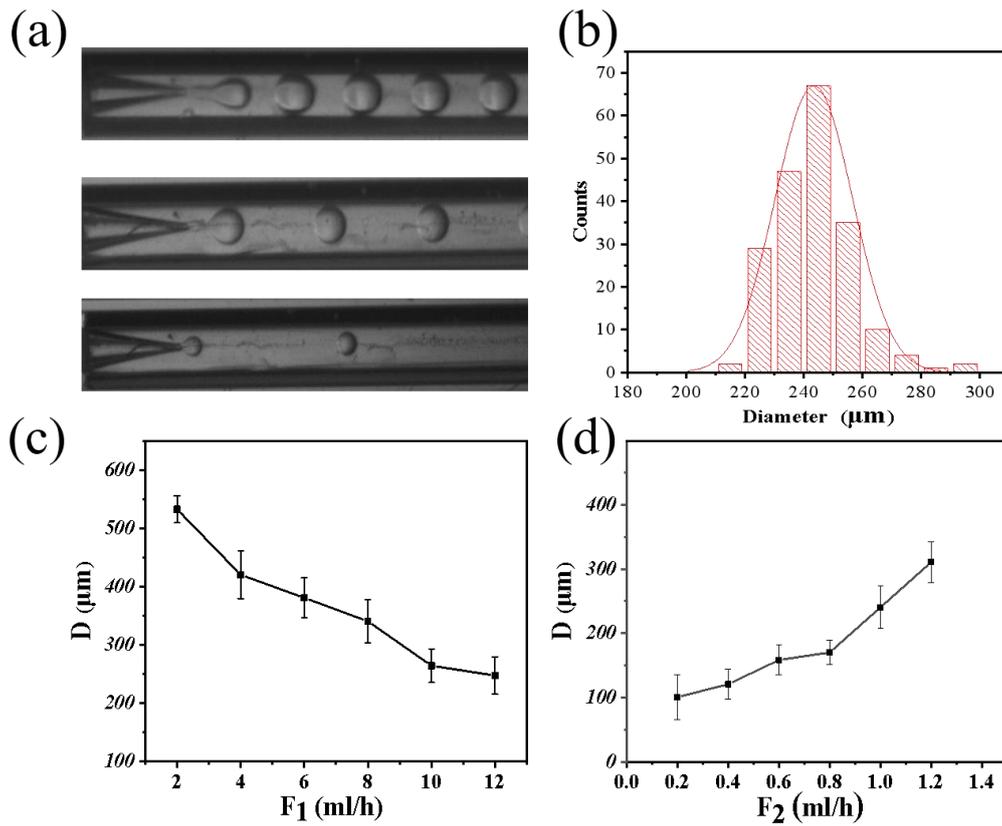
## Supporting Information

### Tailoring esophageal tumor spheroids on a chip with inverse opal scaffolds for drug screening

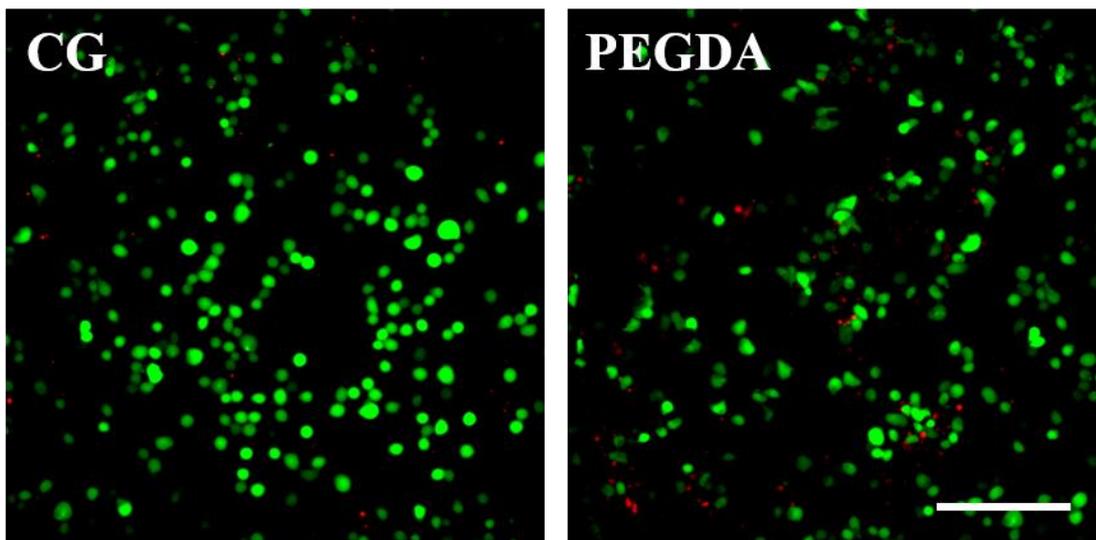
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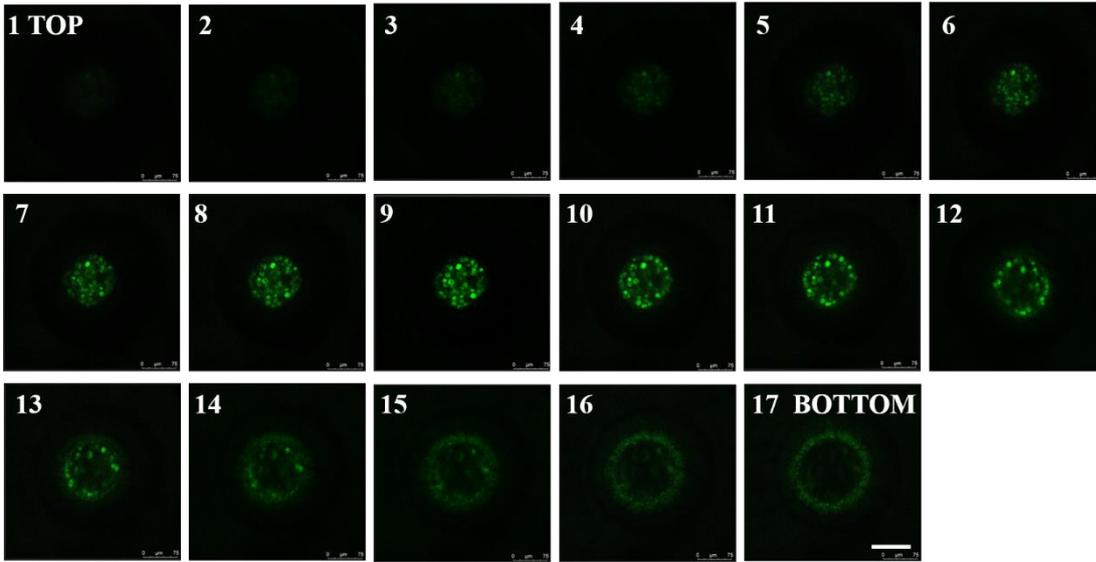
**Figure S1.** Hydrogel molding experiments with different concentrations (20%, 40%, 60%, 80% from left to right). a: Moment of solidification; b: After natural withering.



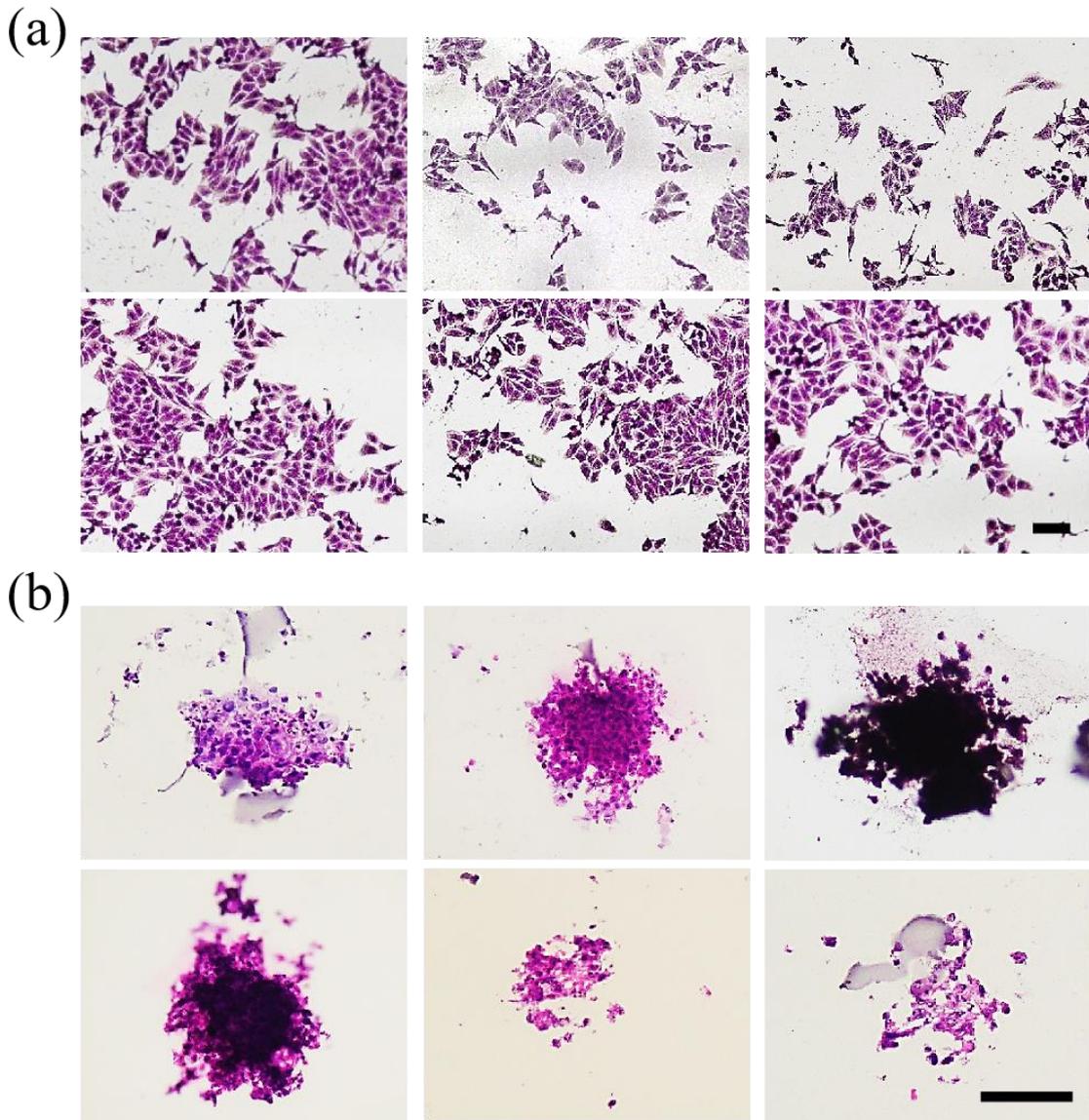
**Figure S2.** (a) Real-time microscopic images of the generation of single emulsion droplets. (b) The size distribution of the droplets. (c) The relationship between the droplet diameter and the inner flow rate ( $F_1$ ). (d) The relationship between the droplet diameter and the outer flow rate ( $F_2$ ).



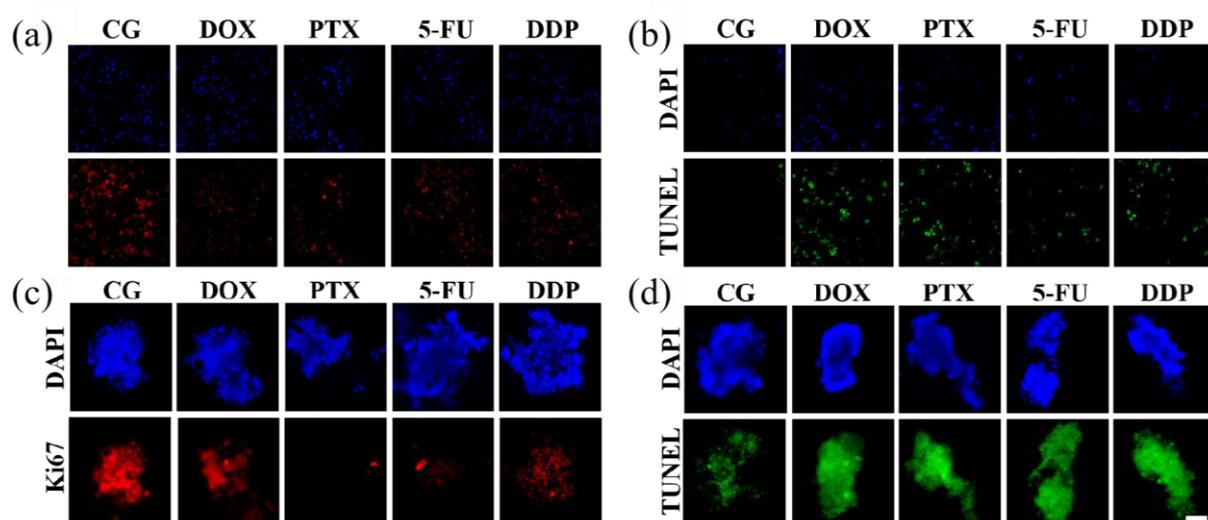
**Figure S3.** Biocompatibility test of materials (green: Calcein AM, red: PI). The scale bar is  $100\mu\text{m}$ .



**Figure S4.** The confocal images of KYSE-70 spheroid in different Z-planes. The scale bar is  $100\mu\text{m}$ .



**Figure S5.** HE staining of esophageal cells. (a) Cell morphology in two-dimensional culture mode. (b) Cell spheroplast morphology in different layers. The scale bar is 100µm.



**Figure S6.** Immunofluorescence images of two-dimensional cell models and three-dimensional spherical polymers. (a) The fluorescence pattern of Ki67 proliferation signal in the two-dimensional model. (b) The fluorescence pattern of TUNEL apoptosis signal in the two-dimensional model. (c) The fluorescence pattern of Ki67 proliferation signal in the three-dimensional model. (d) The fluorescence pattern of TUNEL apoptosis signal in the three-dimensional model. The scale bar is 100 $\mu$ m.